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Quantitative data on the impact of resistance genes on the population dynamics of *P. infestans*

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1 **Abstract**

2 The purpose of AMIGA DL 8.4 is to deliver quantitative data on the selective impact of host
3 resistance genes on the population dynamics of *P. infestans*. Selection pressure causes
4 undesirable shifts in the pathogen population, e.g. towards virulence for specific
5 (combinations of) R-gene(s). Presence of selection pressure indicates the critical need for
6 continuous pathogen population monitoring and a dynamic control strategy to counteract
7 the negative effects of selection pressure.

8 Significant differences between the local pathogen populations in Carlow (Ireland) and
9 Valthermond (the Netherlands) were found: the Carlow population is relatively stable over
10 years and primarily made up of a few well known clonal lines whereas the Valthermond
11 population is highly dynamic over years, contains greater genetic variation and is primarily
12 made up of genetically distinct genotypes plus contributions from clonal line EU_13_A2. The
13 difference in genetic variation contained in both populations is very likely due to the effect
14 of an active sexual cycle in Valthermond.

15 As expected, resistant cultivar Sarpo Mira and resistant, Desiree derived, GM potato clone
16 A15-031 both exerted significant selection pressure on both local pathogen populations. In
17 Carlow, Sarpo Mira was infected by a limited number of specific sub-variants of clonal line
18 EU_13_A2. In Valthermond, Sarpo Mira was also infected by sub-variants of EU_13_A2 plus a
19 range of genetically distinct isolates. Vnt1 containing potato clones were only infected in
20 Valthermond by a range of genetically distinct pathogen genotypes outside the well-known
21 clonal lines. Populations harbouring high levels of genetic variation are thus more likely to
22 overcome R genes. This also illustrates the high potential for rapid change towards a virulent
23 pathogen population once the *Rpi-Vnt1* is used on larger acreages.

24 Investigation of specific selection pressure exerted by GM crops versus non-GM crops
25 remained inconclusive. Insufficient samples were found on the conventional Vnt1 clone,
26 factors “year” and “origin” were intertwined and the pathogen population in Valthermond
27 demonstrated a large background variability between years.

28 It is concluded that cultivation of (GM) resistant potato exerts significant selection pressure
29 on the local *P. infestans* population. To prevent dramatic population shifts towards
30 virulence, rendering host resistance all but useless, population monitoring and a dynamic

1 control strategy must be in place to counteract the negative effects of selection pressure.

2

1 Introduction

2 Potato late blight, one of the world's most devastating plant diseases in potato and tomato,
3 is caused by the oomycete *Phytophthora infestans*. In the past, potato late blight led to e.g.
4 the Great Irish - and Continental Famine (Zadoks, 2008). Currently potato late blight still is
5 the most important disease in potato cultivation, traditionally controlled by highly frequent
6 (calendar based) fungicide applications (Cooke et al., 2011) supported by preventative
7 cultural measures such as crop rotation, the use of healthy seeds and the timely destruction
8 of primary sources of inoculum. Nevertheless potato late blight remains responsible for an
9 estimated annual economic loss of M€ 1000 on the 6 Mha of potato grown in the EU
10 (Haverkort et al., 2008).

11 Host resistance and subsequent cultivation of potato late blight resistant potato cultivars is
12 the most (cost) effective and environmentally friendly way to control potato late blight
13 (Schepers et al., 2009). Currently however, cultivation of resistant cultivars is very limited
14 due to an overwhelming demand for a limited number of commercially successful but highly
15 late blight susceptible cultivars. In addition, potato breeding is complex and time consuming
16 (Rietman et al., 2012) mostly due to the tetraploid nature of the crop. It was also repeatedly
17 shown that resistance gene (R gene) mediated host resistance was easily overcome by the
18 highly adaptive pathogen *P. infestans* (e.g. Black et al., 1953, Fry 2008; Haas et al. 2009;
19 McDonald and Linde 2002), especially if resistance is based on a single R gene. The origin of
20 the adaptive capability of *P. infestans* was shown to reside in the *P. infestans* genome in
21 combination with its high reproductive capacity (Haas & Kamoun et al., 2009). As a result,
22 adaptation is "the *Phytophthora infestans* way of life" resulting in R genes being overcome
23 (e.g. Black et al., 1953, Fry 2008; Haas et al. 2009; McDonald and Linde, 2002), resistance to
24 active ingredients of fungicides (e.g. Dowley and O'Sullivan, 1981) and increased
25 aggressiveness (e.g. Flier and Turkensteen, 1999). The net results are the often dramatic and
26 sudden population changes such as those described by Drenth et al. (1993), Cooke et al.
27 (2012) and Fry et al. (2013).

28 Recently developed GM technology overcomes some of the disadvantages of conventional
29 breeding with respect to breeding for resistance to potato late blight: Stacking R genes in
30 popular commercial potato varieties in a fraction of the time required for conventional
31 breeding while the cultivar characteristics remain the same was successfully demonstrated

1 within the DuRPh project (Haverkort et al., 2016).

2 It is however to be expected that (large scale) cultivation of (GM) resistant potato cultivars
3 leads to selection pressure on the pathogen population for virulent genotypes. Within the
4 DuRPh project a tailor-made potato late blight control strategy based on resistant cultivars
5 aimed to mitigate the effects of selection pressure using a low input fungicide spray program
6 (Haverkort et al., 2016). This strategy was also successfully deployed within the AMIGA
7 project (AMIGA deliverables 8.3 and 8.6). For this purpose, five large scale field trials,
8 containing a susceptible potato cultivar Desiree and two resistant potato cultivars/clones,
9 were carried out in Ireland and the Netherlands. In order to quantify selection pressure, *P.*
10 *infestans* was sampled in and around these field trials. Infections in the field trials were of
11 natural origin and therefore represent the local *P. infestans* population. *P. infestans* sampled
12 in the fields was genetically characterised using the standardized 12-plex Euroblight SSR set
13 (Li et al., 2013). The resulting genotypic data were analysed for impact of resistance on
14 composition of the local *P. infestans* population.

15

1 **Materials and Methods**

2

3 **Field trials**

4 Field trials are fully described in AMIGA DL 8.3 but they are summarized below:

5 Five field trials were carried out, two in the Netherlands in 2013 and 2014 (Valthermond,
6 GPS coordinates 52.873828°, 6.942644) and three in Ireland in 2013, 2014 and 2015 (Oak
7 Park, Carlow, GPS coordinates; 52.8560667, -6.9121167). Trials were carried out under
8 permit IM10-006 for the Netherlands and in Ireland the trials were licensed by the
9 Environmental Protection Agency as per Notification No. B/IE/12/01.

10 Two commercial potato cultivars and one potato clone were used in the field trials: the
11 conventionally bred cultivar Desiree (highly susceptible to potato late blight), the
12 conventionally bred cultivar Sarpo Mira (highly resistant to potato late blight) and the
13 “Desiree based”, cis-genically modified (Jacobsen and Schouten, 2007), highly resistant, *Rpi-*
14 *Vnt1.1* containing clone A15-031 (described in detail in Haesaert et al., 2015). Sarpo Mira is
15 reported to contain the R3a, R3b, R4, *Rpi-Smira1* and *Rpi-Smira2* potato late blight
16 resistance genes (Rietman et al., 2012).

17 IPM field trials were laid out as randomized block experiments including the three potato
18 genotypes mentioned above, three potato late blight control strategies (unsprayed control,
19 weekly spray schedule and a “next level IPM control strategy”, (see AMIGA DL8.3)) and
20 seven replicates.

21 Monitoring plots, located outside the trials themselves, served the purpose of monitoring
22 the local *P. infestans* population for adaptation against the R-genes deployed in the field trial
23 and thus quantification of selection pressure. Trials in Valthermond were surrounded by ten
24 monitoring plots whereas the trials at Oak Park were surrounded by 11 monitoring plots in
25 2014 and 2015. Monitoring was not possible for Ireland in 2013 due to the unavailability of
26 seed of the conventional clone RH06-975-8 containing *Rpi-vnt1.3*. Each monitoring plot
27 contained 6 (NL) or 3 (IE) plants of Desiree, Sarpo Mira and the conventional clone RH06-
28 975-8. RH06-975-8 was used as a conventional substitute for the Cisgenic *Rpi-Vnt1.1*
29 containing clone A15-031. Monitoring plots were not sprayed with crop protection products

1 but at times (e.g. Ireland 2014) received additional irrigation. From emergence onwards,
2 these plots were monitored for infection on a weekly basis. When infection was found, this
3 was quantified as the number of lesions and the percentage destroyed foliage (severity).
4 Also a *P. infestans* DNA sample was taken by pressing the sporulating part of a single lesion
5 on an FTA card (Whatman International Ltd, Maidstone, United Kingdom). A maximum of 4
6 DNA samples was taken per potato genotype, monitoring plot and year. Infections occurring
7 in the field trials were quantified and sampled in the same manner as done for the
8 monitoring plots. However, lesion counts were omitted as it was found to be impractical.
9 FTA cards were collected and stored at room temperature until further molecular analysis.

10

11 **Genotypic characterisation**

12 The Euroblight standardized set of twelve microsatellite markers was used for genotypic
13 characterisation of the *P. infestans* samples (Knapova and Gisi 2002; Lees et al. 2006; Li et al.
14 2010; Li et al. 2013). Amplification of the SSR markers was carried out as described by Li et
15 al. (2010). Capillary electrophoresis was carried out on an automated ABI-3130xl sequencer
16 (Ireland 2014+2015) using a 16-capillary array (36 cm) or an automated 48-capillary array (36
17 cm) ABI 3730 according to the manufacturer's instructions. SSR allele scoring and sizing was
18 done using GeneMapper version 3.7 (Applied Biosystems).

19

20 **Data analysis**

21 For the purpose of this analysis, results from the monitoring plots and the field trials were
22 pooled since no monitoring plot samples were available from the Irish field site. The genetic
23 distance between field samples was determined and visualized using the R package POPPR
24 2.2.1 (Kamvar et al., 2014, Kamvar et al., 2015). SSR data was formatted as a GenAEx csv file
25 in Excel 2010 (© 2010 Microsoft Corporation) and then imported into R as a genclone object.
26 Genetic distances were estimated using Bruvo distance (Bruvo et al., 2004). Subsequently a
27 minimal spanning network (msn) was drawn based on these bruvo distance estimates. Field
28 samples were assigned to groups based on the year, country of origin and host genotype
29 they were sampled from. Each group was assigned a unique colour with similar colours for

1 the same host-genotypes irrespective of country of origin. Sub-setting of data was
2 performed in order to investigate selection pressure from the resistant potato genotypes on
3 *P. infestans* genotypes found. Nodes represent the number of samples with a minimum of 1.
4 Larger nodes therefore represent multiple field samples with identical multi-locus genotypes
5 (mlg). The number inside the node indicates the number of samples represented. The
6 correlation between node size and number of represented samples has been modified by
7 base logarithm 1.3 in order to prevent the larger nodes obscuring results in surrounding
8 nodes. For the field samples from The Netherlands a genetic distance cut-off of 0.3 was
9 used. This means isolates sharing less than 70% genetic similarity (based on SSR alleles) will
10 not be connected in the minimal spanning network. For Ireland a cut-off of 0.1 was set,
11 forcing samples sharing less than 90% genetic similarity into different groups, highlighting
12 groups of samples with a similar clonal mlg. The strict cut-off value for Ireland allows to
13 assert the clonal nature of the population. The higher cut-off value for the Netherlands
14 allows to investigate whether groups of mlg's e.g. share a certain virulence profile. Within
15 the msn, genetic distance is correlated with line thickness: wide black lines indicate lower
16 genetic distance whereas thin grey lines indicate greater genetic distance. The maximum
17 value for genetic distance connected by lines is set by the cut-off value discussed above.

18

1 **Results**

2 From the five field trials, a total of 360 samples were taken, genotypically characterised and
 3 analysed. The origin of these samples is given in Table 1. Irish samples were only sampled
 4 from the AMIGA trial itself.

5 **Country / location effects**

6 Table 1 shows *P. infestans* was not found on Vnt1 containing plant material in Ireland. This
 7 in contrast to the Netherlands where *P. infestans* was found both on A15-031 in the field
 8 trial and on the conventional Vnt1 clone in the monitoring plots. In addition, Figure 1
 9 demonstrates a much higher level of genetic variation present in the *P. infestans* population
 10 in Valthermond (the NL) as compared to the *P. infestans* population in Carlow (Ireland). The
 11 pathogen population at Carlow is primarily made up of three well known clonal lineages:
 12 EU_13_A2 (Blue13), EU_6_A1 (Pink6) and EU_8_A1. The pathogen population at
 13 Valthermond is primarily made up of individuals outside the well-known clonal lineages
 14 apart from 16 samples that likely belong to the EU_13_A2 clonal lineage. The *P. infestans*
 15 populations in both countries are therefore genetically very different.

16 **Table 1.** Origin of *P. infestans* DNA samples taken in and around field trials during the AMIGA project.

Country	Year	Type field	Potato genotype	Nr of DNA samples obtained	Total per year	Total per location
Ireland	2013	Trial	Desiree	48	92	
			Vnt1 clone	0		
			Sarpo Mira	44		
	2014	Trial	Desiree	45	62	
			Vnt1 clone	0		
			Sarpo Mira	17		
	2015	Trial	Desiree	43	43	197
			Vnt1 clone	0		
			Sarpo Mira	0		
Netherlands	2013	Monitoring	Desiree	27	117	
			Vnt1 clone	0		
			Sarpo Mira	7		
		Trial	A15-31	34		
			Desiree	37		
			Sarpo Mira	12		
	2014	Monitoring	Desiree	29	46	163
			Vnt1 clone	7		
			Sarpo Mira	10		

17

1 From Figure 1 it can be concluded that almost all the different *P. infestans* genotypes found
 2 are capable of infecting susceptible cultivar Desiree (light and dark green in Figure 1).
 3 However, only specific *P. infestans* mlgs (clonal variants for IE) are capable of infecting Sarpo
 4 Mira (orange and yellow) and/or Vnt1 containing plant material (shades of red).
 5 Interestingly, for samples from Ireland it was found that only certain sub-variants of
 6 EU_8_A1 were found on Sarpo Mira plants. This could indicate that Sarpo Mira exerts
 7 selection pressure against specific EU_8_A1 isolates.

8 Year Effects

9 **Carlow:** 2013 and 2015 show a more or less equal distribution of clonal lines EU_8_A1 and
 10 E_13_A2 on Desiree (Figure 2). In 2014 the population was mostly dominated by EU_8_A1
 11 with a small contribution from EU_13_A2. The limited number of EU_6_A1 was only found
 12 (on Desiree) in 2014.

13 **Valthermond:** *P. infestans* populations on all three potato cultivars for 2013 and 2014 are
 14 related but different with very few overlapping genotypes between years and a relatively
 15 small contribution of the well-known clonal lineages (Figure 2). Based on the results on
 16 Desiree, the 2014 population seems to be slightly more “clonal” as compared to the 2013
 17 population.

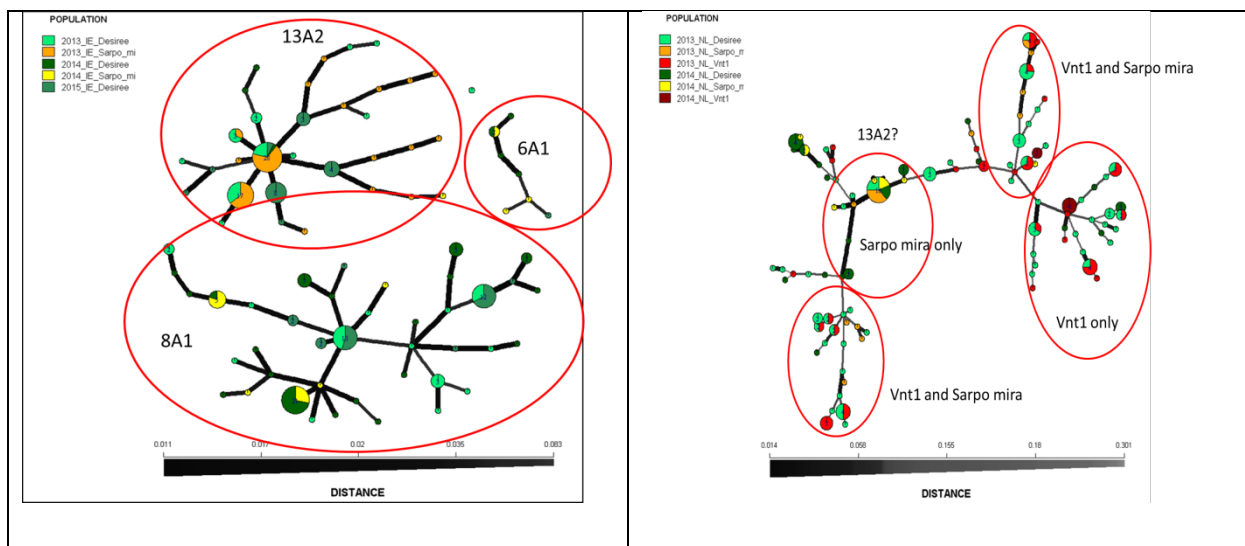
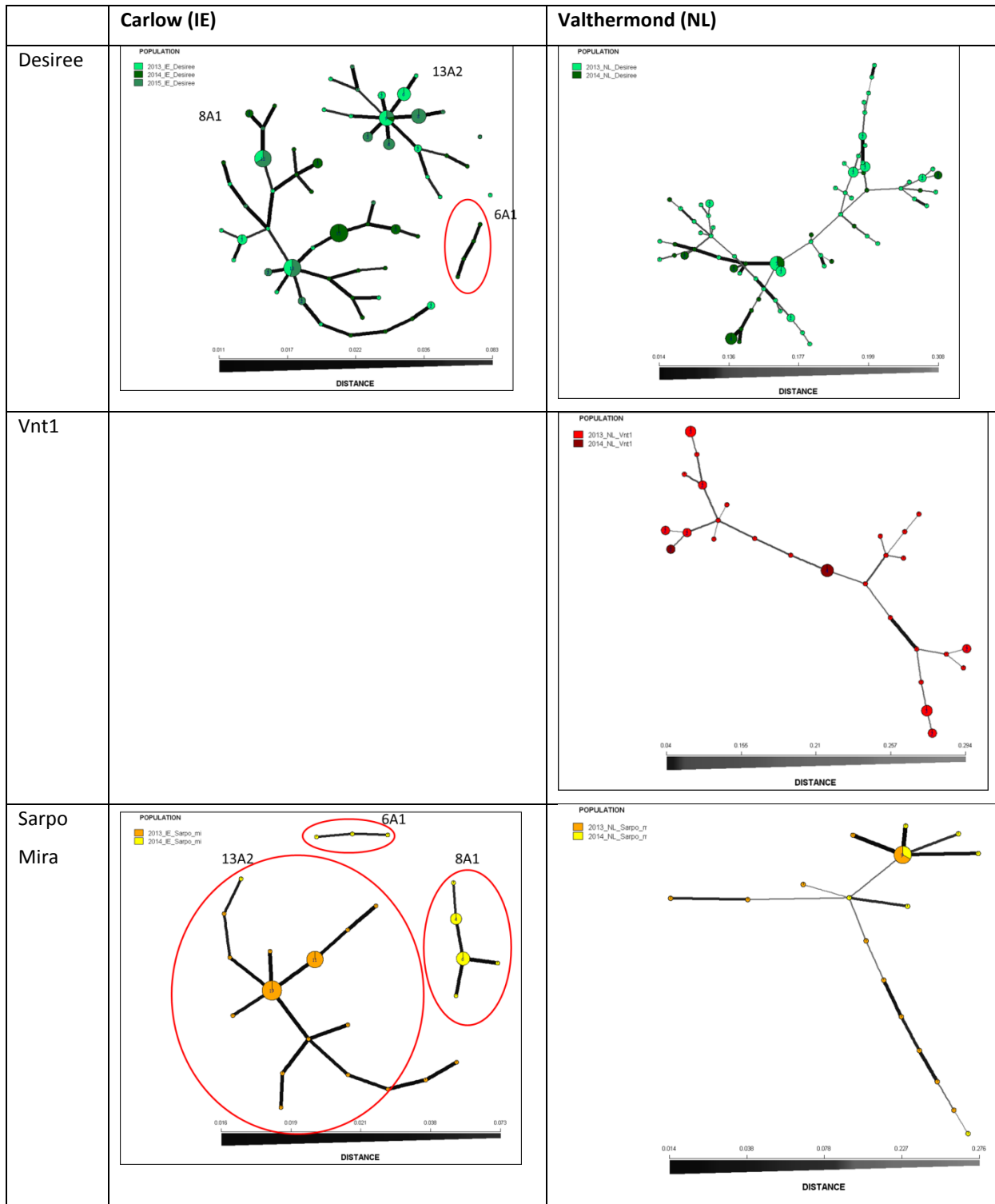


Figure 1. Genetic distance between samples from Ireland (left panel) taken in 2013, 2014 and 2015 and The Netherlands (right panel) taken in 2013 and 2014 as an indication for the genetic variation among *P. infestans* samples in and around the two AMIGA field trials. Numbers in each node indicate the number of identical SSR multi-locus genotypes represented. Distance indicates the genetic distance represented by the lines connecting nodes in the network.



1 **Figure 2.** Genetic variation per potato clone, location and year. Numbers in each node indicate the number of
2 identical SSR multi-locus genotypes represented. Distance indicates the genetic distance represented by the
3 lines connecting nodes in the network.

4

5

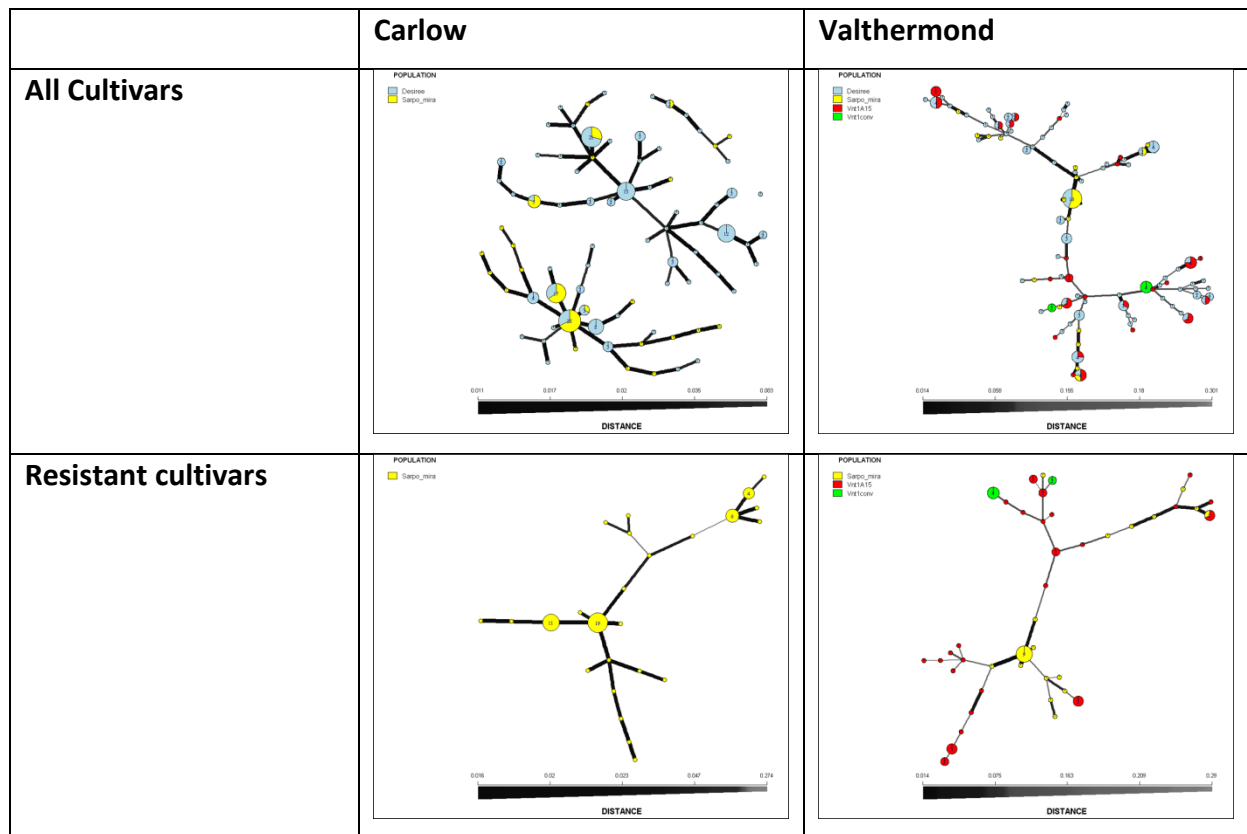
1 **Cultivar effects**

2 **Desiree** : At both locations, Desiree is infected by almost all *P. infestans* genotypes (in blue,
3 top row in Figure 3), demonstrating a good representation of almost all genotypes found or
4 on this susceptible cultivar.

5 **Sarpo Mira**: In Carlow, this cultivar is infected by sub-variants of the well-known clonal lines
6 (EU_13_A2, EU_8_A1 and EU_6_A1). In Valthermond EU_13_A2 variants also infect Sarpo
7 Mira. In addition other, more genetically distinct, *P. infestans* genotypes also occasionally
8 infect Sarpo Mira in Valthermond.

9 **Vnt1 containing plant material**: Vnt1 containing plant material is only infected in
10 Valthermond, the Netherlands (Figure 3). This is very likely due to the large genetic variation
11 contained in the local pathogen population as a result of an active sexual cycle and regular
12 oospore contributions to main-crop infections. Most Vnt1 infecting *P. infestans* genotypes
13 are more genetically distinct and outside the well-known clonal lines described earlier. If we
14 include the results from Figure 1, Vnt1 infecting *P. infestans* genotypes can be grouped in at
15 least three groups, with or without the additional capability to infect Sarpo Mira. Based on
16 the data presented it can be concluded that virulence against Vnt1 can be found amongst a
17 wide range of genetically distinct *P. infestans* genotypes. It is therefore likely that this
18 ability/virulence can spread quickly throughout sexually recombining *P. infestans*
19 populations such as those in The Netherlands. In contrast none of the traditional clonal
20 multi-locus genotypes were found on Vnt1 containing plants.

21



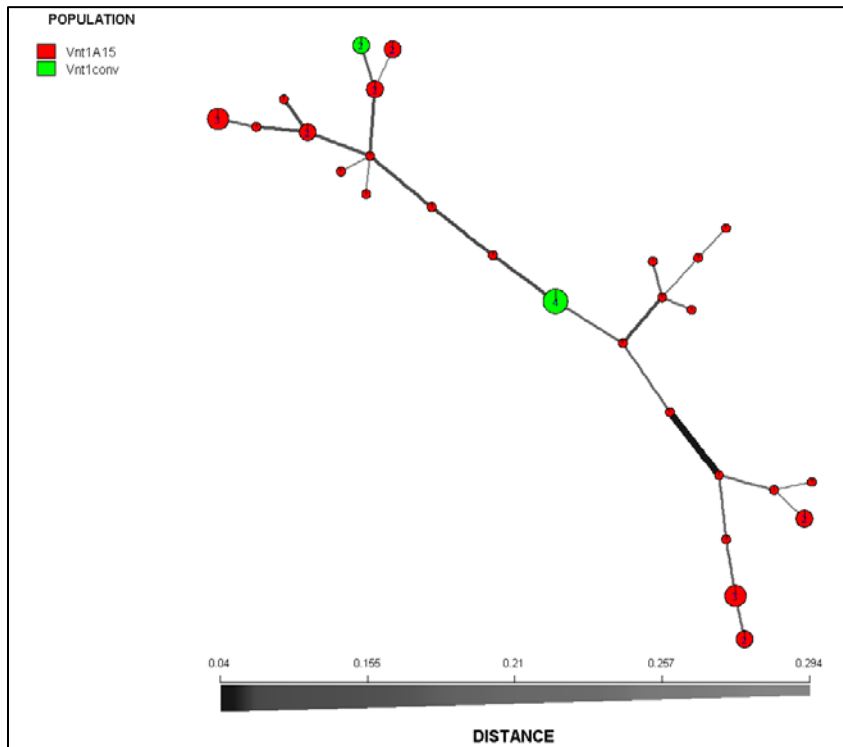
1 **Figure 3.** Genetic variation of the local *P. infestans* population on Desiree, Sarpo Mira and Vnt1 containing plant
2 material. Numbers in each node indicate the number of identical SSR multi-locus genotypes represented.
3 Distance indicates the genetic distance represented by the lines connecting nodes in the network.

4

5 **Conventional Vnt1 versus GM Vnt1 containing plant material**

6 Figure 4 gives the genetic distances for *P. infestans* samples taken from the cisgenic *Rpi-*
7 *Vnt1.1* containing potato line A15-031 and *P. infestans* samples taken from conventional *Rpi-*
8 *Vnt1.3* containing potato clone RH06-975-8. Despite the fact that *Rpi-Vnt1.1* and *Rpi-Vnt1.3*
9 are reported to share the same resistance spectrum (Pel et al., 2009), conventional-Vnt1
10 infecting *P. infestans* genotypes are genetically distinct from GM-Vnt1 infecting *P. infestans*
11 (green- versus red nodes in Figure 4). Also within both groups only a few identical mlgs are
12 found. This is a direct indication for selection pressure in general and selection of different *P.*
13 *infestans* genotypes by the GM and non-GM, Vnt1 containing potato clones. All 34 samples
14 found on A15-031 are however from 2013 whereas all 7 samples from the conventional Vnt1
15 clone are from 2014. In this case, the “year factor” is thus confounded with the factor
16 “genotype of origin” resulting in an inconclusive answer on the question whether a
17 genetically modified potato selects for distinct *P. infestans* genotypes when compared to

1 conventionally bred potato varieties. This is the case even more if we consider the
2 differences between the 2013 and 2014 *P. infestans* populations in Valthermond reported
3 above. Last, but not least, this finding might also be an indication of the presence of other R
4 genes in the conventional Vnt1 containing potato clone.



5
6 **Figure 4.** Genetic distance between *P. infestans* samples taken from the cisgenic potato line A15-031 (red)
7 compared to *P. infestans* samples taken from conventional potato clone Vnt1 (green). Numbers in each node
8 indicate the number of identical SSR multi-locus genotypes represented. Distance indicates the genetic
9 distance represented by the lines connecting nodes in the network.

10
11

1 Discussion

2 The purpose of AMIGA DL 8.4 is to deliver quantitative data on the impact of host resistance
3 genes on the population dynamics of *P. infestans*. Selection pressure causes shifts in the
4 pathogen population towards e.g. virulence for specific (combinations of) R-gene(s).
5 Occurrence of selection pressure indicates the critical need for continuous pathogen
6 population monitoring and a dynamic control strategy to counteract any negative effects of
7 selection pressure.

8 For this purpose *P. infestans* was sampled and genetically characterised in and around the
9 field trials performed in Ireland and the Netherlands. The results (Figure 1) indicate clear
10 differences between the local pathogen populations in Carlow (Ireland) and Valthermond
11 (the Netherlands). The Carlow population is primarily made up of a few well known clonal
12 lines whereas the Valthermond population contains greater genetic variation and is primarily
13 made up of genetically distinct isolates plus contributions from clonal line EU_13_A2. Over
14 the three years, the composition of the Carlow population was more or less stable with
15 varying contributions of the same clonal lines. In Valthermond, EU_13_A2 was present in all
16 three years but the majority of the pathogen population was genetically distinct between
17 both years (Figure 2). The difference in genetic variation contained in both populations is
18 very likely due to the effect of an active sexual cycle in the *P. infestans* populations in or
19 around the Valthermond site. The highly undesirable result, as illustrated, is that R genes
20 and thus host resistance in general is more likely to be overcome by pathogen populations
21 harbouring high(er) levels of genetic variation.

22 Susceptible cultivar Desiree hosted almost all of the *P. infestans* genotypes found in Carlow
23 and Valthermond. Resistant cultivar Sarpo Mira and the resistant, Desiree based, cisgenically
24 derived A15-031 both projected significant selection pressure on the pathogen population
25 (e.g. Figure 3). In Carlow, Sarpo Mira was infected by a limited number of sub-variants of
26 clonal line EU_13_A2, EU_6_A1 and EU_8_A1. In Valthermond, Sarpo Mira was also infected
27 by sub-variants of EU_13_A2 plus a range of genetically distinct isolates. Vnt1 containing
28 potato clones were not infected in Carlow which is consistent with published evidence that
29 EU_13_A2 does not overcome the Vnt1 gene (Cooke et al., 2012). In Valthermond they were
30 infected by a range of genetically distinct pathogen genotypes indicating the potential for
31 rapid changes towards a virulent pathogen population once the R gene is used on larger

1 acreages.

2 An attempt to investigate specific selection pressure exerted by GM crops versus non-GM
3 crops remained inconclusive. Insufficient samples were successfully genotyped on the
4 conventional Vnt1 clone, factors “year” and “origin” were confounded: all conventional Vnt1
5 samples came from 2014 whereas all GM Vnt1 samples came from 2013 and the pathogen
6 populations in Valthermond demonstrate a large variability between years. Last, but not
7 least, this finding might also be an indication of the presence of other R genes in the
8 conventional Vnt1 containing potato clone.

9 Overall, it is clear that cultivation of resistant potato exerts significant selection pressure on
10 local *P. infestans* populations. To prevent selection driven, dramatic pathogen population
11 shifts, rendering host resistance all but useless, population monitoring and a dynamic
12 control strategy must be in place to counteract the negative effects of selection pressure.
13 Host resistance breaking traits would need to be countered by careful deployment of
14 resistance genes in space and time (Skelsey et al 2010) in combination with a low input spray
15 strategy when necessary (AMIGA DL 8.3 + 8.6).

16

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4

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